amplification of a genomic fragment. A similar strategy is used to screen for lines homozygous for the 35S-GFP T-DNA. First, the presence of the 35S-GFP T-DNA is demonstrated by using the T-DNA-specific PCR primer LB1 and the gene-specific PCR primer L22F4F (5'- ttc gaa aac att acc tcc gat c-3', SEQ ID NO:30). Second, plants carrying the 35S-GFP T-DNA are tested for homozygosity by using the gene-specific primers L22F4F and F22L4R (5'-ggc ttt tgc att tgg tat cta cta g-3', SEQ ID NO:31) The plants homozygous for both the S11.13-34 and 8Z-2 transgenes and plants homozygous for the 8Z-2 transgene but with no S11.13-34 transgene are allowed to self fertilize to obtain F3 generation plants. These plants and the parental line 8Z-2 are scored for incidence of PTGS based on GFP fluorescence as described in Example 4. The results summarized in Table 3 show that PTGS of the 35S-GFP transgene is lost in plants with a T-DNA insert interrupting the region encoding a polypeptide comprising an RNase D-related domain.--

REMARKS

Page 52, Example 5 has been amended to correct an obvious typographical error in a sentence that describes plant lines carrying the T-DNA are checked for homozygosity. One skilled in the art would understand that if the lines were already known to be homozygous for the T-DNA, then there would be no need to check for homozygosity for the T-DNA. It is obvious that the lines resulting from the crosses and known to have a T-DNA insert are checked for homozygosity.

No new matter has been added by this amendment.

No fee is believed due with this amendment. If any fees are required, please charge them to the Deposit Account No. 50-1744 (in the name of Syngenta).

Respectfully submitted,

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